

# OneStep RTPCR Kit

Cat.No. ODR61

**Description:** The OneStep RTPCR Kit contains all necessary reagents for both reverse transcription and PCR amplification, using gene-specific primers and target RNAs from either total RNA or mRNA.

*Taq* PCR Smart Mix (2X) is a proprietary mix of *Taq* DNA polymerase, Buffer, dNTPs and  $MgCl_2$  for easy and convenient use. The product is recommended to use amplification less than 5kb PCR product.

## Product contents

Contents	Volume
RTase (200U/ $\mu$ L)	5000U
5X RT Reaction Buffer (Included 50mM DTT mix)	200 $\mu$ L
10mM dNTP	100 $\mu$ L
RNase-free Water	600 $\mu$ L
Random hexamer (2.8nmol)	Lyophilized Dissolve in 56 $\mu$ L nuclease free water.
Oligo(dT) <sub>18</sub> (2.8nmol)	Lyophilized Dissolve in 56 $\mu$ L nuclease free water.
<i>Taq</i> PCR Smart Mix (2X) (without dye)	1mL

**Note:** Prepare aliquots of Random Hexamer and Oligo (dT)<sub>18</sub> to avoid multiple freeze thaw cycles.

## Storage

- Store at  $-20^{\circ}C$

## Features

Features	
RNase H activity	RNase H negative
Temperature of cDNA synthesis	At $42-55^{\circ}C$
Synthesis length	< 14 kb
Synthesis Primer	Random primer, Oligo (dT) <sub>18</sub> , Gene specific

## Protocol

- **First Strand cDNA synthesis (Total reaction volume - 20 $\mu$ L)**

1. Prepare the following mixture in a PCR tube.

RNA	10ng – 1 $\mu$ g total RNA 10ng – 0.5 $\mu$ g mRNA	- $\mu$ g
Primer	1 $\mu$ L Oligo (dT) <sub>18</sub> / 1 $\mu$ L Random hexamer or 0.5 $\mu$ L Oligo (dT) <sub>18</sub> + 0.5 $\mu$ L Random hexamer or 1 $\mu$ L Gene-specific Primer (15 – 20 $\mu$ M)	
dNTP (10mM)		1 $\mu$ L
Make up the volume with RNase-free Water to		15.5 $\mu$ L

2. Heat the mixture at 65°C for 5 minutes and cool down immediately on ice for 1 minute. Spin down the tube by centrifugation briefly.

**Note: Use heating block or PCR machine. DO NOT use water bath.**

3. Add the following components to the same tube:  
(Total reaction volume - 20 $\mu$ L)

Reagents	Volume
5X RT Reaction Buffer	4 $\mu$ L
RTase	0.5 $\mu$ L

4. Mix gently.

**Note:** For random hexamer, incubate the combined reaction mixture at 25°C for 5 minutes and then proceed with the next step. For oligo (dT)<sub>18</sub> or gene-specific primers, directly proceed with the next step.

5. Incubate at 50°C for 10 minutes.

6. Inactivate the reaction mixture by heating at 95°C for 5 minutes.

**Note: Use heating block or PCR machine. DO NOT use water bath.**

7. The product can be used for PCR immediately or stored at -20°C.

**Note: As a recommendation; cDNA product should compose 10% of total reaction volume of PCR.**

- **PCR amplification using cDNA as template**

Recommended PCR mixture and cycling condition

PCR mixture (Reaction vol. 50 $\mu$ L)		Cycle		
<i>Taq</i> PCR Smart Mix (2X)	25 $\mu$ L	95°C	2 min	×1
Forward primer (10pmol/ $\mu$ L)	2 $\mu$ L	95°C	20 sec	} ×35-40
Reverse primer (10pmol/ $\mu$ L)	2 $\mu$ L	AT	40 sec	
Template cDNA (<200ng)	- $\mu$ L	72°C	1 min/kb	
Add D.W to	50 $\mu$ L	72°C	5 min	×1

Modifications on the amount of template, extension time, annealing temperature, and the number of PCR cycles can be done according to the target size, primer's  $T_m$ , and the type of templates for amplification.